

Preliminary communication

Sialic acids of a new type from the lipopolysaccharides of *Pseudomonas aeruginosa* and *Shigella boydii*

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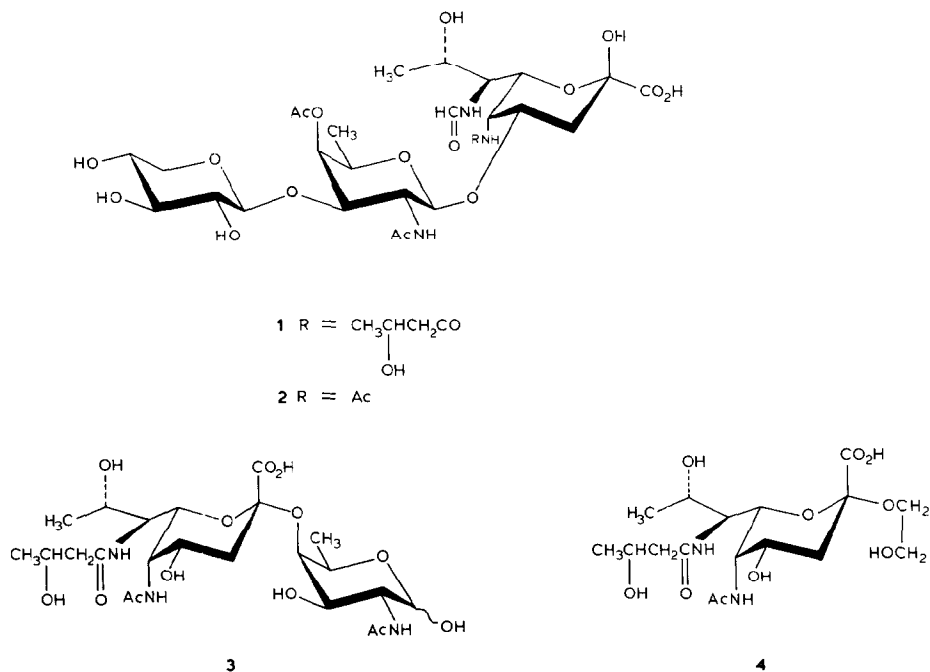
All sialic acids hitherto found in Nature are derivatives of 5-amino-3,5-dideoxy-D-glycero-D-galacto-nonulosonic (neuraminic) acid¹. We now report the identification of sialic acids of a new type, in lipopolysaccharides from *P. aeruginosa* O5 and O10 (Lányi classification²) and *Sh. boydii* type 7, as derivatives of 5,7-diamino-3,5,7,9-tetra-deoxynonulosonic acid (pseudaminic acid). The new sialic acids were not isolated in a free state and their structures were established by ¹H- and ¹³C-n.m.r. spectroscopy of oligosaccharides derived from the lipopolysaccharides.

The lipopolysaccharides, isolated from dry bacterial cells of *P. aeruginosa* O5a,b,c and O5a,b,d by the Westphal procedure³, reacted with a resorcinol reagent to give a chromophore identical to that formed from *N*-acetylneuraminic acid⁴. In accordance with the earlier observation⁵, mild acid hydrolysis (1% CH₃CO₂H, 100°, 1.5 h) of the lipopolysaccharides degraded the O-specific polysaccharide chains, and oligosaccharides 1 and 2 were subsequently isolated by gel filtration on Sephadex G-50 and were acidic in electrophoresis. On solvolysis with HF (20°, 3 h), the oligosaccharides yielded D-xylose and 2-acetamido-2,6-dideoxy-D-galactose[†], which were identified by conventional methods, an acidic component being undetected.

The ¹H- and ¹³C-n.m.r. data showed that 1 and 2 were trisaccharides containing xylose, 2-acetamido-2,6-dideoxygalactose, and a 3-deoxyaldulosonic acid [δ_C^* 99.7 (s, C-2) and 34.8 (t, C-3); δ_H 2.07 (dd, $J_{3e,4}$ 5, $J_{3e,3a}$ 12 Hz, H-3e) and 1.91 (t, $J_{3a,4}$ 12 Hz, H-3a)]. Unambiguous assignment of all signals in the ¹H-n.m.r. spectra of 1 and 2 was performed by homonuclear double-resonance, and all signals in their ¹³C-n.m.r. spectra were then assigned by selective heteronuclear ¹³C{¹H} double-resonance. The data obtained indicated that the xylose and 2-acetamido-2,6-dideoxygalactose residues in 2 were β -linked [δ_C^* 105.4 and 99.6 (J_{CH} 162 and 163 Hz, respectively⁷, C-1), δ_H 4.37 and 4.72 (2 d, each $J_{1,2}$ 8 Hz, H-1)], and thus the aldulosonic acid was the reducing residue. The chemical shifts for xylose in the ¹³C-n.m.r. spectrum of 2 were very similar to those reported for methyl β -D-xylopyranoside⁸; therefore, the xylosyl group occupied the terminal position.

[†]Previously, these sugars were found to be the components of serologically related *P. aeruginosa* types 7 and 8 (Habs classification)⁵ as well as immunotype 6 (Fischer)⁶ lipopolysaccharides.

[‡]The δ_C^* data refer to gated-decoupling spectra.



Further, the 2-acetamido-2,6-dideoxygalactosyl residue was substituted by xylose at position 3 [δ_C 78.9 (C-3)] and by an O-acetyl group at position 4 [δ_H 5.24 (H-4)].

The 1H and ^{13}C chemical shifts for the aldulosonic acid in **2** indicated it to be a diaminotetradexonulosonic acid, the amino groups being attached to positions 5 and 7 (δ_C 46.6 and 52.3) and two deoxy groups occupying positions 3 (see above) and 9 [δ_C 16.3; δ_H 1.10 (d, 3 H, $J_{8,9}$ 6 Hz, H-9)]. One of the amino groups of the sialic acid residue in **2** was acetylated (δ_C 23.5, δ_H 1.98, CH_3 CON), whereas the other carried a formyl group [δ_C^* 164.9 (d, J_{CH} 197 Hz); δ_H 8.05 (s)]. The new sugar was substituted by 2-acetamido-2,6-dideoxygalactose at O-4 (δ_C 72.3, C-4), whereas HO-8 was unsubstituted (δ_C 67.5, C-8).

Comparison of the ^{13}C -n.m.r. spectra of **1** and **2** indicated that they differed only by the presence of an *N*-(3-hydroxybutyryl) group on the sialic acid residue in **1** [δ_C 65.9 (CHOH), 45.5 (t, CH_2), 23.5 (CH_3); δ_H 2.21 (d, J 6 Hz, CH_2) and 1.39 (d, J 6 Hz, CH_3)] instead of an *N*-acetyl group in **2**. The change of the *N*-acyl substituent caused distinct shifts of the signals for C-1 of the 2-acetamido-2,6-dideoxygalactosyl and C-4 of the sialic acid residues, from 99.6 and 72.3 p.p.m. in **2** to 100.6 and 73.2 p.p.m., respectively, in **1**, whereas the position of all other signals remained unaltered. The above shifts were indicative of the location of the substituent near the glycosidic linkage between these residues, *i.e.*, at C-5 of sialic acid, and thus the formamido group was attached to C-7.

Analysis of the ^{13}C -n.m.r. spectra of the corresponding lipopolysaccharides showed that trisaccharides **1** and **2** represent their chemical repeating-units and that the sialic acid residues in **1** and **2** existed almost completely in the same anomeric form as in the lipopolysaccharides.

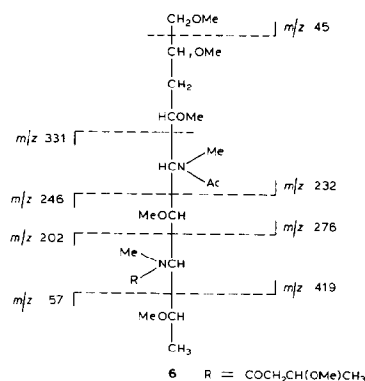
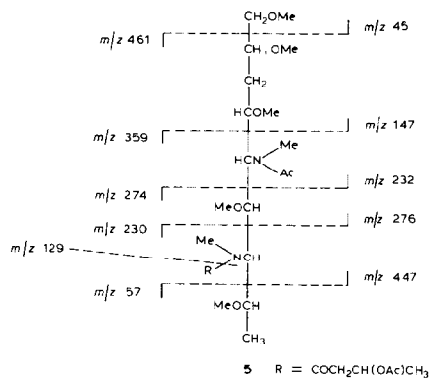
Lipopolysaccharides from *P. aeruginosa* O10a and *Sh. boydii* type 7 also contained an unusual sialic acid, but, in contrast to the above lipopolysaccharides, they gave the corresponding polysaccharides on hydrolysis, which were isolated by gel filtration on

Sephadex G-50. The results of acid hydrolysis, solvolysis with HF, and ^1H - and ^{13}C -n.m.r. spectroscopy proved in *P. aeruginosa* O10a polysaccharide to be composed of trisaccharide repeating-units made up of 2-acetamido-2,6-dideoxy-D-glucose, 2-acetamido-2,6-dideoxy-D-galactose, and an *N*-acetyl-*N'*-(3-hydroxybutyryl)diaminotetradexynonulosonic acid, while the *Sh. boydii* type 7 polysaccharide possesses a pentasaccharide repeating-unit containing D-glucose, D-galactose, 2-acetamido-2-deoxy-D-glucose, and the same sialic acid, in the ratios 1:2:1:1.

Solvolysis of *P. aeruginosa* O10a polysaccharide with HF (20°, 3 h) yielded *N*-acetyl-6-deoxyhexosamines and an oligosaccharide **3** (isolated by gel filtration on Sephadex G-15). Assignment of the signals in the ^1H - and ^{13}C -n.m.r. spectra (as above) showed that **3** was a disaccharide, the 2-acetamido-2,6-dideoxygalactose residue being at the reducing end (the spectra contained two series of signals corresponding to the α and β forms) and the sialic acid occupying the terminal position [δ_{C}^* 102.6 (s, C-2)]. The ^{13}C chemical shifts for the sialic acid in **3** (δ_{C} 37.1, 67.7, 49.1, 75.0, 54.7, 69.3, and 17.7 for C-3,4,5,6,7,8,9) showed the same distribution of functional groups as for the sialic acid residues in **1** and **2**, while the similarity of the ^1H coupling constants ($J_{3e,4}$ 5, $J_{3a,4}$ 12, $J_{4,5}$ <1, $J_{5,6}$ 1.5, $J_{6,7}$ 10, $J_{7,8}$ 6, and $J_{8,9}$ 6 Hz) proved the configuration of the sialic acid residues in **1**, **2**, and **3** to be the same.

Some differences in chemical shifts of the signals for C-3,4,5,7,8 of the sialic acid residues in **2** and **3** were associated with glycosidation at C-4 in **2** and replacement of the formyl group at N-7 in **2** by an acetyl or 3-hydroxybutyryl group in **3**. The relatively high-field position of the signal for C-6 of the sialic acid in **2** (71.2 p.p.m.), as compared to that (75.0 p.p.m.) in **3**, is characteristic of a different anomeric configuration of these sugars (axial orientation of the carboxyl group in **3** and equatorial in **2**⁹). This conclusion was also supported by the chemical shifts for H-3e in the ^1H -n.m.r. spectra of **3** (δ 2.55) and **2** (δ 2.07)¹⁰.

Sh. boydii type 7 polysaccharide was subjected to three successive Smith-degradations to give glycoside **4** which, according to the ^{13}C -n.m.r. data, was composed of sialic acid [δ_{C}^* 101.8 (s, C-2)] and ethylene glycol as the aglycon (δ_{C} 66.8 and 61.8). Similar chemical shifts for C-3,4,5,6,7,8,9 in the ^{13}C -n.m.r. spectra of **3** and **4**, as well as the similarity of the ^1H coupling constants for the acetylated derivatives of **3** and **4**, showed that these oligosaccharides contained the same sialic acid. This finding is in agreement with the strong serological cross-reaction¹¹ between *P. aeruginosa* O10a and *Sh. boydii* type 7.



The position of the *N*-acyl substituents, as well as the whole structure of the sialic acid residues, was evident from the mass spectra of the derivatives 5 and 6, which were prepared as follows. *P. aeruginosa* O10a and *Sh. boydii* type 7 polysaccharides were carboxyl-reduced¹² and then hydrolysed (0.01 M HCL, 100°, 2 h) to split the glycosidic linkage of the sialic acid residue, and the products were reduced with sodium borohydride. The oligosaccharide thus obtained from the former polysaccharide was methylated¹³ and then solvolysed with HF, and the product was acetylated to give 5. The oligosaccharide derived from the latter polysaccharide was solvolysed with HF and the product was methylated¹³ to form 6. The *O*-acetylation of the *N*-(3-hydroxybutyryl) group in 5 proved it to be glycosylated in the corresponding polysaccharide, whereas the sialic acid residue was unsubstituted.

Thus, the new sialic acids are derivatives of the same 5,7-diamino-3,5,7,9-tetra-deoxynonulosonic (pseudaminic) acid. Determination of the configuration of pseudaminic acid (tentatively identified as *L-glycero-L-manno*) and of the 3-hydroxybutyryl residue is in progress.

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